

Protective Role of Flavonoid on Histological Architecture of Kidney and Enzymatic Functions of Liver in Heat-Stressed Male Mice

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Abstract: The current study of the evaluation of renal and hepatic function of male mice exposed to the stressed environment condition. In the experiment design, twenty-four male mice have been divided into three groups: the control group drenched normal saline, the stressed group drenched normal saline and kept for 3 weeks, and the flavonoid group drenched flavonoid 25 mg/kg b.w. for 3 weeks. After 24 hours of flavonoid administration, synthesized mice with kitamine were subjected for blood draining to measurement serum MDA, GPx, AST, and ALT as well as to collection the kidney specimens for histology. The result of serum antioxidant enzyme MDA and GPx among groups of experiment showed highly significant effect between stressed group under heat stress and control. The decrement of serum MDA in the flavonoid group showed a significant effect when compared with the stressed group, while there was a slightly significant difference between the control and flavonoid groups showed significant decreased serum GPx of stressed group, while increase in serum GPx of flavonoid group were seen compared to stressed group, and slightly significant between control and flavonoid group. The result of hepatic function referred to effect of stressed on liver significant elevation in serum AST and ALT of stressed in comparison to other groups. Histopathological effects of stressed environment on renal tissues showed influx of the inflammatory cells to the renal tissues and epithelial degeneration appears. Also, hemorrhage, edema, fatty degeneration and accumulation of inflammatory cells were observed. Flavonoid administration to the adult male mice showed decreased effects of stressed on renal tissues through reduction inflammatory cells, edema, hemorrhage and epithelial cell degeneration when compared with the stressed group. In conclusion, the experiment referred to the effects of stressed environmental conditions on cellular activity through effects on cellular enzymes, cell membrane regulation, and metabolism.

Keywords: Antioxidants, Environmental conditions, Malondialdehyde, Glutathione peroxidase, Liver function, Iraq.

Introduction

Stressed environmental conditions have various impacts to tissues functions of different body organs through an imbalance in the antioxidant systems glutathione and MDA and upregulation of the hepatic enzymes ALT and AST. These results refer to the downregulation of the antioxidant system and the upregulation of serum hepatic enzymes (Chen et al., 2023; Ding et al., 2023; Hu et al., 2023). Stressed environmental conditions cause liver injury, which is the most common complication in animals and leads to disturbances in liver function and metabolism due to increased cellular reactive oxygen species (ROS) that disrupt cell enzyme function and cellular oxidative stress. These mechanisms refer to effects on mitochondrial function and disturbances in the cellular antioxidant system and apoptotic pathway that cause apoptosis and autophagy in hepatic tissues showing elevation to pro-inflammatory cytokines and lipid peroxidation (Cheng et al., 2019; Emami et al., 2020; Ji et al., 2024; Kabiri-Arani et al., 2024; Rahmani et al., 2024). Stressed environment causes heat shock protein stimulation to protect cellular damage from oxidative stress, on the other hand, the relationship between cellular metabolism energy and hypothalamus pituitary adrenal axis to regulation of cellular environment refers to the role of oxidative stress through enhanced reactive oxygen species and inflammatory responses that cause cellular hepatic damage (Balakrishnan et al., 2023; Kizir et al., 2024; Madkour et al., 2024; Yang et al., 2024).

Renal function regulation of the cellular plasma in the body, a stressed environment leads to decreased renal plasma due to decreased renal blood flow through stimulation of the renin angiotensin aldosterone system (RAS); these mechanisms cause vasoconstriction of renal vessels (Hansson et al., 2020; Wesseling et al., 2020; Chapman et al., 2021; Rebez et al., 2023). Stressed environments affect cellular function through increased free radicals resulting from cellular oxidative stress, which leads to downregulation of the antioxidant system and upregulation of the apoptotic pathway, leading to renal cell apoptosis and enhanced cytokine levels with inflammatory reactions causing renal damage (Imbabi et al., 2023; Ke et al., 2024; Wang et al., 2024). In addition, the effects of heat stress on blood platelets due to a reduction plasma fibrinogen and platelet aggregation (Ke et al., 2024). Histopathological analysis of renal tissues showed cellular influx of neutrophils, basophils, and lymphocytes; cellular aggregation led to edema, hemorrhage, and inflammation. Beside this, fatty degeneration of renal epithelial tubules and chronic exposure to heat stress lead to renal failure caused by necrosis and degeneration of renal tubules (Chen et al., 2020; Chapman et al 2021).

Methodology

The experiment has been conducted healthy twenty four male mice weighted 34 ± 2 gm aged 2 month. A total of 24 adult male mice were randomly divided into three equal groups (8 mice/ group) were intubated daily by using gavage and handled for three weeks. Control group were intubated normal saline and kept at 22 ± 1 , heat-stressed group were drenched normal saline and kept at 32 ± 1 for 3 weeks, flavonoid group were intubated flavonoid 25 mg/kg b.w. daily and kept at 32 ± 1 for 3 weeks. After end of experiment animals synthesized with kitamine 100 mg/kg b.w. and xylazine 10 mg/kg b.w. serum collected from centrifuged blood to estimate serum (MDA, GPx, ALT, and AST) and took kidney sample for histopathological examination, and kept at 10% formalin.

Statistical analysis

The result of the experiment was analyzed by graph pad prism program compression performed using one-way ANOVA to detect significant differences between values of study groups at $p < 0.05$ (Gharban, 2023).

Result

The result of serum antioxidant enzyme MDA and GPx among groups of experiment showed highly significant ($p < 0.05$) effect between stressed group under heat stress and control. The decrement of serum MDA in flavonoid group showed a significant ($p < 0.05$) effect when compared with the stressed group, while there was a slightly significant ($p < 0.05$) difference between the control and Flavonoid groups figure 1. In the figure 2 showed significant decreased serum GPx of stressed group when compared with control, while significant increase of serum GPx of flavonoid group when compared with stressed group, and slightly significant between control and flavonoid group. The result of hepatic function referred to effect of stressed on liver (Figures 3 and 4) significant elevation in serum AST and ALT in stressed group compared to others. Histopathological effects of stressed environment on renal tissues (Figure 5) showed influx of the inflammatory cells to the renal tissues and epithelial degeneration appears compared to others (Figure 5C) so that, hemorrhage, edema, fatty degeneration and accumulation of inflammatory cells when compared to control. Flavonoid administration 25 mg/kg b.w. to the adult male mice showed decreased effects of stressed on renal tissues through reduction inflammatory cells, edema, hemorrhage and epithelial cell degeneration compared to stress group (Figure 5 Q).

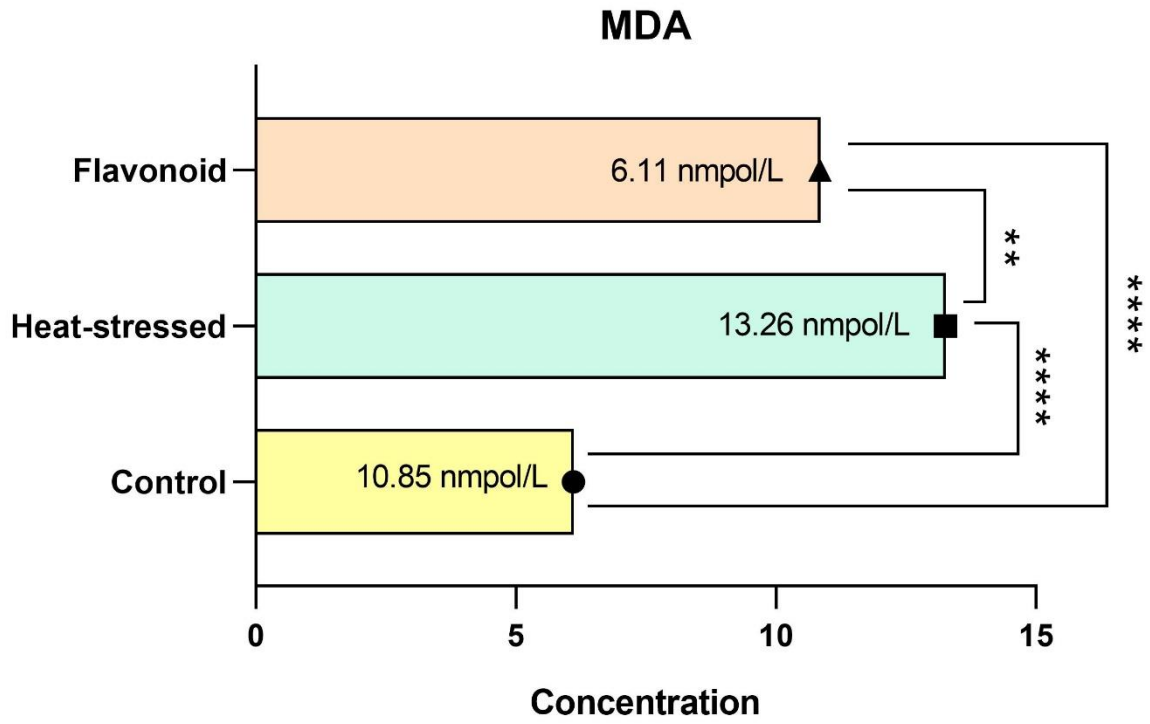


Figure 1: Effect of stress on serum MDA

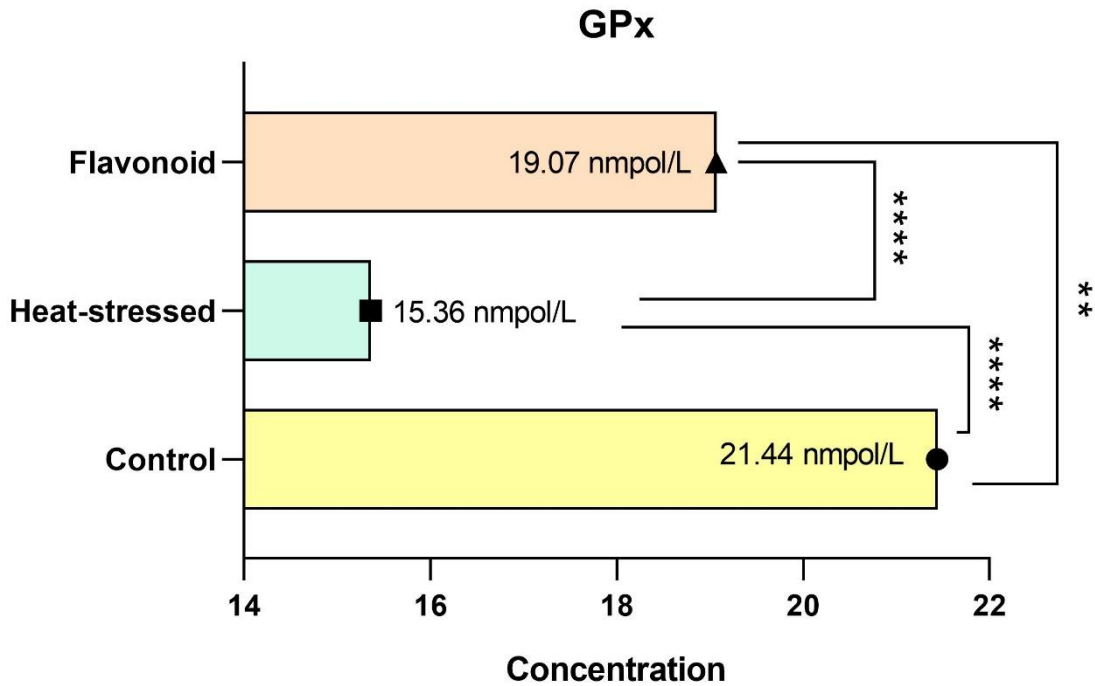


Figure 2: Effect of stress on serum GPx

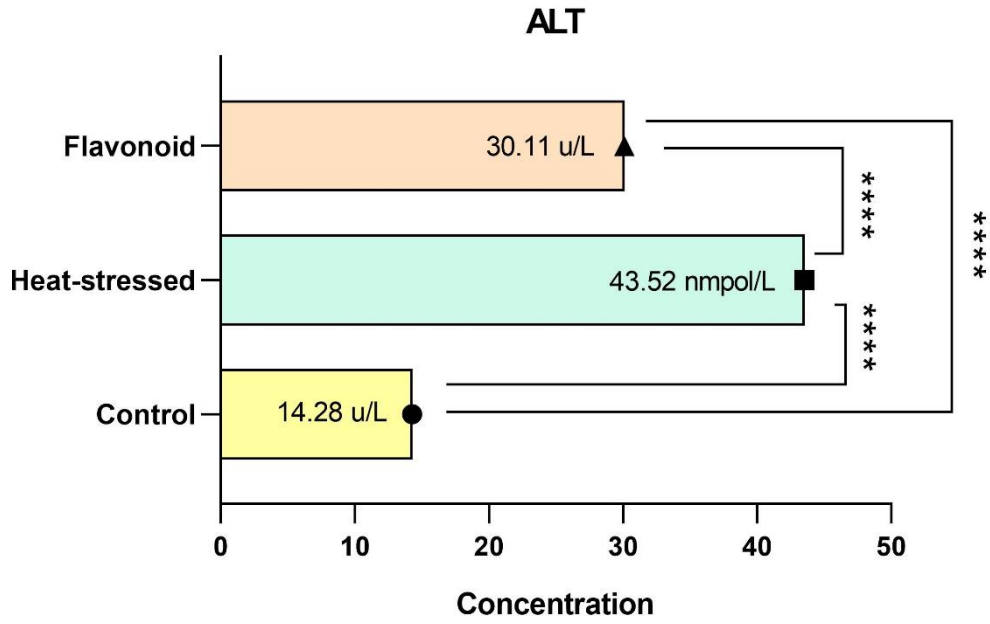


Figure 3: Effect of stress on serum ALT

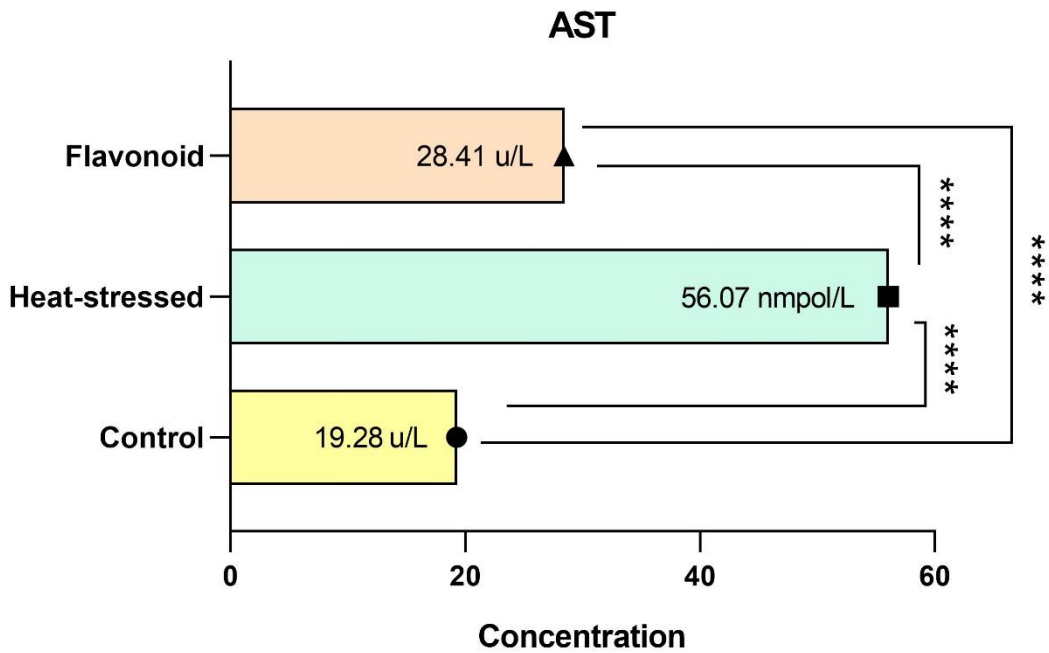


Figure 4: Effect of stress on serum AST

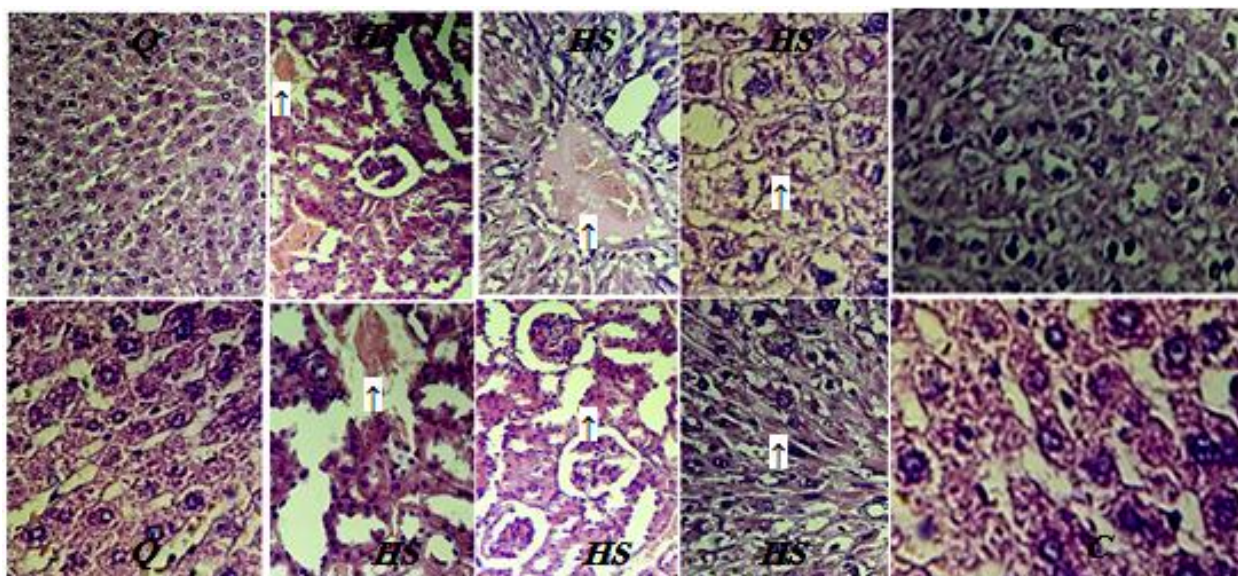


Figure 2: Renal histology in study mice

Discussion

The result of the experiment confirm to the effects of stressed environmental condition on the physiological function of liver and kidney, the result of serum antioxidant record effects of heat stress on lipid peroxidation due to significant increase of serum MDA in stressed group when compared with the control group, while significant decrease in serum GPx when compared with the control group, these result referred to the effects of stressed environment condition on the antioxidant system and apoptotic pathway lead to influx of free radicals causes cellular oxidative stress through enhancement in reactive oxygen species, these material effect on mitochondrial metabolism regulation and causes apoptosis to the renal and hepatic cells (Malyar, et al., 2020; Delkhosh, et al., 2021; Aderao, et al., 2023; Chinko, and Umeh, 2023; Mbepera, et al., 2023; He, et al., 2024; Li, et al., 2024). So, the study showed the effects of stress on the function of hepatic and renal cells through an increase in serum ALT and AST. The result referred to a significant effect of the stressed group when compared with the control group of serum ALT and AST. These findings confirm the role of the stressed environment on cellular function through a downregulation of antioxidants, an upregulation of lipid peroxidation, and an increase in free radicals and oxygen species that effect cell membrane function and lead to the efflux of enzymes out of the cell (Malyar et al., 2021; Zhang et al., 2022; Ding et al., 2023; Rahmani et al., 2024; Yang et al., 2024).

On the other hand, the result of the experiment recorded the role of Flavonoid on cellular function regulation to decrease the effect of oxidative stress and free radicals result from a stressed environment through downregulation of lipid peroxidation reduction serum MDA and upregulation of glutathione peroxidase. These mechanisms ensure the effect of Flavonoid on cellular mechanism through a decrease oxygen species and free radicals that cause disturbances in cellular function. The experiment result was a decrease in serum MDA of the Flavonoid treatment group when compared with the stressed group and an increase in serum GPx of the Flavonoid group when compared with the stressed group. The result referred to the administration of Flavonoid as an antioxidant and cellular protective role (Alsharidah et al., 2021 ; Khalifa et al., 2021; Almatroodi et al., 2021; Ghonim, 2023; Abd El-Hamid et al., 2024; Manoharan et al., 2024). In addition, the role of Flavonoid treatment in regulating cellular activity through regulation of ALT and AST cellular enzymes refers to the effect of Flavonoid on cellular function and enzyme regulation through reduction of free radicals and oxygen species that disrupt cell membrane function. The experiment result showed a significant effect of Flavonoid on oxidative stress through downregulation of serum ALT and AST when compared with the stressed group. These results confirm the effects of flavonoid on cellular mechanism regulation and protection from oxidative stress and reactive oxygen species (Ghonim et al., 2023; Caglar et al., 2024; Hafez et al., 2024; Raghunandhakumar et al., 2024). The result of the renal histopathologic section showed

effects of heat stress on renal tissue through a significant influx of inflammatory cells, hemorrhage, edema, and degeneration of tubule epithelial cells. These referred to disrupting balance between the antioxidant system and oxidants that lead to increased lipid peroxidation, free radicals, and reactive oxygen species that cause disturbances in the apoptotic pathway that resulted in the degeneration and accumulation of inflammatory cells (Hansson et al., 2020; Khalil et al., 2020; Chapman et al., 2021). Administration of Flavonoid 25 mg/kg b.w. daily for three weeks reduced the effects of heat stress on renal tissues through decreased inflammatory cells and degeneration when compared with the control group. These mechanisms referred to the effects of Flavonoid on regulation of the cellular antioxidant system and the apoptotic pathway that decreased inflammatory cell influx and degeneration (Alsharidah et al., 2021; Ashour et al., 2021).

Conclusion

The result of the experiment referred to the effects of stressed environmental conditions on cellular activity through effects on cellular enzymes, cell membrane regulation, and metabolism. That results in the effect of heat stress on lipid peroxidation, hepatic function, and renal function through effects on serum MDA, GPX, ALT, and AST, so it effects on histopathological examination of renal tissues influx of inflammatory cells and degeneration, while administration of flavonoid reduces the effects of the stressed environment condition through stimulating the antioxidant system and regulating the apoptotic pathway, leading to decreased lipid peroxidation and degeneration through reduction of inflammatory cells.

References

1. Abd El-Hamid, M. I., El-Azzouny, M. M., El-Malt, R. M., Elkenawy, M. E., Abdelwarith, A. A., Younis, E. M., and Ibrahim, D. (2024). Future impact of Flavonoid-loaded nanoemulsion in rabbits: prospects for enhancing growth, immunity, antioxidant potential and resistance against *Pasteurella multocida*. *Frontiers in Veterinary Science*, *10*, 1340964.
2. Aderao, G. N., Jadhav, S. E., Pattanaik, A. K., Gupta, S. K., Ramakrishnan, S., Loksha, E., and Singh, G. (2023). Dietary selenium levels modulates antioxidant, cytokine and immune response and selenoproteins mRNA expression in rats under heat stress condition. *Journal of Trace Elements in Medicine and Biology*, *75*, 127105.
3. Almatroodi, S. A., Alnuqaydan, A. M., Alsahli, M. A., Khan, A. A., and Rahmani, A. H. (2021). Flavonoid, the most prominent constituent of *Nigella sativa*, attenuates liver damage in streptozotocin-induced diabetic rats via regulation of oxidative stress, inflammation and cyclooxygenase-2 protein expression. *Applied Sciences*, *11*(7), 3223.
4. Alsharidah, M., Abdel-Moneim, A. M. H., Alsharidah, A. S., Mobark, M. A., Rahmani, A. H., Shata, A., and Al Rugaie, O. (2021). Flavonoid, but not metformin, protects against gentamicin-induced nephrotoxicity and renal dysfunction in rats. *Applied Sciences*, *11*(9), 3981.
5. Ashour, H., Rashed, L., Elkordy, M. A., Abdelwahed, O. M., ASHOUR, H., RASHED, L., and ABDELWAHED, O. (2021). Flavonoid ameliorates acute kidney injury induced by renal ischemia-reperfusion. *International Journal of Morphology*, *39*(2), 469-476
6. Balakrishnan, K. N., Ramiah, S. K., and Zulkifli, I. (2023). Heat shock protein response to stress in poultry: A review. *Animals*, *13*(2), 317.
7. Caglar, K., Dokuyucu, R., Agturk, G., Tumer, C., Tutuk, O., Gocmen, H. D., and Gogebakan, B. (2024). Effect of Flavonoid on transient receptor potential melastatin (TRPM) channels in rats with liver ischemia reperfusion model in rats. *Iranian Journal of Basic Medical Sciences*, *27*(3), 319.
8. Chapman, C. L., Johnson, B. D., Parker, M. D., Hostler, D., Pryor, R. R., and Schlader, Z. (2021). Kidney physiology and pathophysiology during heat stress and the modification by exercise, dehydration, heat acclimation and aging. *Temperature*, *8*(2), 108-159.
9. Chapman, C. L., Johnson, B. D., Parker, M. D., Hostler, D., Pryor, R. R., and Schlader, Z. (2021).

Kidney physiology and pathophysiology during heat stress and the modification by exercise, dehydration, heat acclimation and aging. *Temperature*, 8(2), 108-159.

10. Chen, B., Yang, B., Zhu, J., Wu, J., Sha, J., Sun, J., and Zhang, X. (2020). Hsp90 relieves heat stress-induced damage in mouse kidneys: Involvement of antiapoptotic PKM2-AKT and autophagic HIF-1 α signaling. *International Journal of Molecular Sciences*, 21(5), 1646.
11. Chen, S., Zhou, J., Igbokwe, C. J., Duan, Y., Cai, M., He, Y., and Zhang, H. (2023). Oligopeptide of RDPEER from watermelon seeds prevents heat stress-induced liver injury by suppressing oxidative stress and inflammation responses. *Journal of Functional Foods*, 105, 105563.
12. Cheng, K., Yan, E., Song, Z., Li, S., Zhang, H., Zhang, L., and Wang, T. (2019). Protective effect of resveratrol against hepatic damage induced by heat stress in a rat model is associated with the regulation of oxidative stress and inflammation. *Journal of thermal biology*, 82, 70-75.
13. Chinko, B. C., and Umeh, O. U. (2023). Alterations in lipid profile and oxidative stress markers following heat stress on wistar rats: Ameliorating role of vitamin C. *Biomed. Sci*, 9, 12-17.
14. Delkhosh, A., Shoorei, H., Niazi, V., Delashoub, M., Gharamaleki, M. N., Ahani-Nahayati, M., and Abbasgholizadeh, F. (2021). Coenzyme Q10 ameliorates inflammation, oxidative stress, and testicular histopathology in rats exposed to heat stress. *Human and experimental toxicology*, 40(1), 3-15.
15. Ding, K. N., Lu, M. H., Guo, Y. N., Liang, S. S., Mou, R. W., He, Y. M., and Tang, L. P. (2023). Resveratrol relieves chronic heat stress-induced liver oxidative damage in broilers by activating the Nrf2-Keap1 signaling pathway. *Ecotoxicology and Environmental Safety*, 249, 114411.
16. Emami, N. K., Jung, U., Voy, B., and Dridi, S. (2020). Radical response: effects of heat stress-induced oxidative stress on lipid metabolism in the avian liver. *Antioxidants*, 10(1), 35.
17. Gharban, H. A. (2023). Molecular prevalence and phylogenetic confirmation of bovine trichomoniasis in aborted cows in Iraq. *Veterinary world*, 16(3), 580-587.
18. Ghonim, A. (2023). Ameliorative effect of Flavonoid against acrylamide-induced hepatotoxicity in male rats. *Benha Veterinary Medical Journal*, 43(2), 31-35.
19. Hafez, M. H., Ez Elarab, S. M., Tohamy, H. G., and El-Far, A. H. (2024). Flavonoid attenuates diabetes-induced hepatic damage in rat via regulation of oxidative/nitrosative stress, apoptosis, and inflammatory cascade with molecular docking approach. *Scientific Reports*, 14(1), 13016.
20. Hansson, E., Glaser, J., Jakobsson, K., Weiss, I., Wesseling, C., Lucas, R. A., and Wegman, D. H. (2020). Pathophysiological mechanisms by which heat stress potentially induces kidney inflammation and chronic kidney disease in sugarcane workers. *Nutrients*, 12(6), 1639.
21. He, Y., Yu, J., Song, Z., Tang, Z., Duan, J. A., Zhu, H., and Cao, Z. (2024). Anti-oxidant effects of herbal residue from Shengxuebao mixture on heat-stressed New Zealand rabbits. *Journal of Thermal Biology*, 119, 103752.
22. Hu, Y., Lin, L., Liu, K., Liu, E., Han, S., Gong, Z., and Xiao, W. (2023). L-Theanine alleviates heat stress-induced impairment of immune function by regulating the p38 MAPK signalling pathway in mice. *Food and Function*, 14(1), 335-343.
23. Imbabi, T. A., Habashy, W. S., Abol-Fetouh, G. M., Labib, M. M., Osman, A., Elkelish, A., and Ahmed-Farid, O. (2023). Enhancing semen quality, brain neurotransmitters, and antioxidant status of rabbits under heat stress by acacia gum, vitamin C, and lycopene as dietary supplements: an in vitro and in silico study. *Italian Journal of Animal Science*, 22(1), 321-336.
24. Ji, R., Chen, J., Xu, J., Zhang, L., Liu, L., and Li, F. (2024). Protective effect of chlorogenic acid on liver injury in heat-stressed meat rabbits. *Journal of Animal Physiology and Animal Nutrition*.
25. Kabiri-Arani, S., Motallebi, M., Taheri, M. A., Kheiripour, N., Ardjmand, A., Aghadavod, E., and

- Shahaboddin, M. E. (2024). The effect of heat-killed lactobacillus plantarum on oxidative stress and liver damage in rats with bile duct ligation-induced hepatic fibrosis. *Probiotics and Antimicrobial Proteins*, 16(1), 196-211.
26. Ke, H. Y., Chen, J. H., Kao, S. Y., Tsao, C. M., Kuo, C. W., Wu, C. C., and Shih, C. C. (2024). Heat stress-induced platelet dysfunction is associated with loss of fibrinogen and is improved by fibrinogen supplementation. *Thrombosis Research*, 109091.
27. Khalifa, A. A., Rashad, R. M., and El-Hadidy, W. F. (2021). Flavonoid protects against cardiac mitochondrial DNA loss, oxidative stress, inflammation and apoptosis in isoproterenol-induced myocardial infarction in rats. *Heliyon*, 7(7).
28. Khalil, S. R., Salem, H. F., Metwally, M. M., Emad, R. M., Elbohi, K. M., and Ali, S. A. (2020). Protective effect of Spirulina platensis against physiological, ultrastructural and cell proliferation damage induced by furan in kidney and liver of rat. *Ecotoxicology and Environmental Safety*, 192, 110256.
29. Kizir, D., Karaman, M., Demir, Y., and Ceylan, H. (2024). Effect of tannic acid on doxorubicin-induced cellular stress: Expression levels of heat shock genes in rat spleen. *Biotechnology and Applied Biochemistry*.
30. Li, T., Wang, L., Feng, Y., Li, S., Wang, M., Zhao, C., and Zhao, W. (2024). Study on the mechanism of enzymatically treated mulberry (*Morus atropurpurea* Roxb.) leaf protein relieves liver injury in heat stress rats. *Journal of Functional Foods*, 119, 106312
31. Madkour, M., Alaqaly, A. M., Soliman, S. S., Ali, S. I., and Aboelazab, O. (2024). Growth performance, blood biochemistry, and mRNA expression of hepatic heat shock proteins of heat-stressed broilers in response to rosemary and oregano extracts. *Journal of Thermal Biology*, 119, 103791.
32. Malyar, R. M., Li, H., Liu, D., Abdulrahim, Y., Farid, R. A., Gan, F., and Chen, X. (2020). Selenium/Zinc-Enriched probiotics improve serum enzyme activity, antioxidant ability, inflammatory factors and related gene expression of Wistar rats inflated under heat stress. *Life sciences*, 248, 117464.
33. Malyar, R. M., Naseri, E., Li, H., Ali, I., Farid, R. A., Liu, D., and Chen, X. (2021). Hepatoprotective effects of selenium-enriched probiotics supplementation on heat-stressed wistar rat through anti-inflammatory and antioxidant effects. *Biological Trace Element Research*, 199, 3445-3456.
34. Manoharan, N., Parasuraman, R., Jayamurali, D., Muthusamy, P., and Govindarajulu, S. (2024). Role of Flavonoid on sleep restriction and its mitigating effect on leptin-mediated signaling pathway in rat brain. *Molecular Biology Reports*, 51(1), 769.
35. Mbepera, S. M., Mshamu, S. A., Max, R. A., and Malago, J. J. (2023). Heat Stress Induces Oxidative Stress and Predisposes Rats to Gestational Diabetes Mellitus. *Journal of Biology and Life Science*, 14(2), 90-102.
36. Raghunandhakumar, S., Ezhilarasan, D., and Shree Harini, K. (2024). Flavonoid protects thioacetamide-induced chronic liver injury by inhibiting TGF- β 1/Smad3 axis in rats. *Journal of Biochemical and Molecular Toxicology*, 38(4), e23694.
37. Rahmani, M. M., Nasim, M., Shinwari, A. W., and Darmel, M. B. (2024). Amelioration of Heat Stress-Induced Alterations in Immune indices, Serum Enzyme Activity, Antioxidant Ability and Gene Expression in Wistar Rats through Nutritional Strategies. *NUIJB*, 113-118.
38. Rebez, E. B., Sejian, V., Silpa, M. V., and Dunshea, F. R. (2023). Heat stress and histopathological changes of vital organs: A novel approach to assess climate resilience in farm animals. *Sustainability*, 15(2), 1242.

39. Wang, J., Wang, K., Shi, X., Hu, Z., Zhao, L., Chen, K., and Liu, C. (2024). Extreme Heat Exposure Induced Acute Kidney Injury through NLRP3 Inflammasome Activation in Mice. *Environment and Health*.
40. Wesseling, C., Glaser, J., Rodríguez-Guzmán, J., Weiss, I., Lucas, R., Peraza, S., and Jakobsson, K. (2020). Chronic kidney disease of non-traditional origin in Mesoamerica: a disease primarily driven by occupational heat stress. *Revista Panamericana de Salud Pública*, 44.
41. Yang, X., Wang, H., Shen, C., Dong, X., Li, J., and Liu, J. (2024). Effects of isorhamnetin on liver injury in heat stroke-affected rats under dry-heat environments via oxidative stress and inflammatory response. *Scientific Reports*, 14(1), 7476.
42. Zhang, X., Jia, Y., Yuan, Z., Wen, Y., Zhang, Y., Ren, J., and Wei, Y. (2022). Sheng Mai San ameliorated heat stress-induced liver injury via regulating energy metabolism and AMPK/Drp1-dependent autophagy process. *Phytomedicine*, 97, 153920.