



## Down-Regulation of *Shank3* Gene in Patients with Autism Spectrum Disorders and its Reflects in Ability to Speech, Education and Social Communication

Mahdi Saber Al-Deresawi, Aseel Razak Al-Rekaabi  
College of Science / University of Wasit

**Abstract:** The aim of the study was to evaluate how the level of the *Shank3* gene expression influenced speech ability, education, and social communication in children with autism spectrum disorders. A total of forty ASD patients admitted in the (Rahman Specialist Centre taking care of and serving autistic children/ Baghdad/Iraq) and ten children in a seemingly healthy control group were selected. Aged 3-10 years of patients and healthy control group. The results relative to sex showed that the number of males increased significantly (77.5%), as compared to that of females (22.5%). The ability to speak was weak among patients with ASD, and the increase (72.5) was very significant compared to the other group. Although the moderately increased ability to education (65.5), and ability to social communication (65.5) were experienced among patients with ASD. The quantitative Real-Time PCR data revealed that there was a significant reduction (0.374). ASD patients compared to the healthy control group ( $1.00 \pm 0.00$ ) = -0.18). The down-regulation of *Shank3* appears a positively correlated to the Ability to speak, the Ability to educate and the Ability to socialize.

**Key words:** Autism Spectrum Disorders, Quantitative Real-Time PCR, *Shank3* gene.

### 1. Introduction:

Numerous sensory impairments, particularly in the area of tactile sensitivity, are indicative of autism spectrum disorders (ASD) [Tomcheck et al., 2007]. Among these, aberrant nociception—which shows up as either hypersensitivity or hyposensitivity to painful stimuli—is remarkably prevalent in ASD. the prevalence of unreported wounds, self-mutilation (including self-extraction of teeth), and self-harm (Moore, 2015). Neonatal hypotonia, moderate to severe intellectual disability, absent to severely delayed speech, and mild dysmorphic characteristics are common in these patients (Phelan and McDermid, 2012). Autism is a lifelong condition that first appears before the age of three. Additionally, synaptic connections between neurons are formed, reinforced, and altered during the first few years of life. In recent years, a large number of genes essential for synaptogenesis and synapse function have been linked to a variety of neurological and psychiatric conditions, including ASD (Mitcheel, 2010). One copy of *Shank3* losing its function is known to induce Phelan-McDermid syndrome (PMS), which in turn causes ASD and intellectual disabilities (Tachibana et al., 2017; Mitz et al., 2018). Specifically, it has been frequently documented that people with ASD have mutations in genes that code for scaffold proteins such as neuroligins, neurexins, and SHANK as well as synaptic cell adhesion molecules (Geschwind, 2009). These proteins are essential for the development and maintenance of synapses (Huguet et al., 2013). Thus, the several genetic alterations that impact chromatin remodeling, synaptic translation, formation, and function have made the synapse a common target (Sudhof, 2008). The scaffold proteins found at the post-synaptic density of glutamatergic synapses are encoded by shank genes (Toro et al., 2010). While SHANK1 causes the expansion of spine heads, Shank2 and *Shank3* positively influence the induction and maturation of dendritic spines in neurons. Autism spectrum diseases have been linked to shank gene mutations



(Leblond et al., 2014). The 57 kb genome of the human *Shank3* gene, which is found on chromosome 22q13.3, has 24 exons. Exon 18, which is mostly found in the brain, is one of seven exons that are alternatively spliced (Leblond et al., 2014). The purpose of this study was to determine how the *Shank3* gene's level of expression affected the speech, social communication, and educational abilities of children with autism spectrum disorders.

## 2. Materials and Methods:

### 2.1. Ethical Approval:

Prior to their inclusion in the study tests, the Iraqi Ministry of Health and the Ethics Committee of the Department of Biology, College of Science, University of Wasit, gave their approval. Every participant in the study had their father's signed written consent secured.

### 2.2. Sample Collection:

Ten children who appeared healthy served as a control group, while 40 ASD patients who were admitted to the Rahman Specialist Centre for the Care and Service of Autistic Children in Baghdad, Iraq, had three milliliters of blood drawn. Patients ranged in age from three to ten, as did the healthy control group. The center's consultant medical staff has made a clinical diagnosis of the illness. For molecular analysis, EDTA tubes have been filled with each obtained blood sample.

### 2.3. Gene Expression:

Total RNA of all samples was extracted using the TRIzol®LS Reagent according to the manufacturer's instructions. Total RNA was reversely transcribed to complementary DNA (cDNA) using WizScript™ RT FDMix Kit. The procedure was carried out in a reaction volume of 20 µl. The reverse transcription step was carried out one cycle using the next program: 25°C for 10 min, 42 °C for 10 min, 85°C for 5 min and 4°C to the end of the run. The expression levels of *Shank3* gene were estimated by quantitative real time-PCR (qRT-PCR). To confirm this expression EVA Green was used. The mRNA levels of reference gene Glyceraldehyde 3-phosphate dehydrogenase (gapdh) were amplified and used to normalize the mRNA levels of the shank gene.

### 2.4. PCR Reaction and Program:

Quantitative Real-Time PCR reaction was performed using specific primers. Lyophilized primers were dissolved in a free DNase/RNase water to give a final concentration of (100 pmol/µl) as stock solution, to prepare 10 µM concentration as work primer resuspended 10 pmol/µl in 90 µl of deionized water to reach a final concentration 10 µM as work solution, the program of the reaction was : Initial denaturation: 95°C for 5 min (on cycle), Denaturation: 95°C for 40 sec, annealing (gapdh =58°C *shank3*=55°C) for 40 sec, Extension :72 °C for 1 min , the run carried out with 35 cycles then holding with 4 °C for 1 cycle). The sequences of gapdh gene primers was F:5'-AACTTTGGCATTG TGGAAGG-3', R:5'-ACACATTGGGGGTAGAACA-3' [9] and *Shank3* gene was (F:5'-CTGCGCTCCAAGTCCATGACA-3,R:5'-GGCCCTGG CGTTCAAACAATG -3'.

### 2.5. Statistical Analysis:

$\Delta CT$  and  $\Delta\Delta CT$  were calculated according to Livak and Schmittgen equation [2001]. The statistical analysis system –SAS program [2004] was used to the effect of difference factors in traits in this study. Least significant difference (LSD) test was used to the significant compare between means.

## 3. Results and Discussions:

### 3.1. Distribution according ASD features:

Distribution of patients according the gender showed there were a significant ( $P\leq 0.05$ ). increase in male (77.5%) compare to female (22.5%). The patient with ASD recorded a weak ability to speech in high significance ( $P\leq 0.01$ ) increase (72.5%) compared to other group. While the ability to education and ability to social communication were moderate in high significant ( $P\leq 0.01$ ) increase (65.5%) in patient with ASD. Table (1)





**Table (1): Distribution of Case according to Ability to Speech, Education and Social communication**

Factor		No	%	P-value
Gender	Male	31	77.50	0.0005 **
	Female	9	22.50	
Ability to speak	Good	1	2.50	0.0001 **
	Moderate	10	25.00	
	Weak	29	72.50	
Ability to Education	Good	6	15.00	0.0001 **
	Moderate	26	65.00	
	Weak	8	20.00	
Ability to Social communication	Good	6	15.00	0.0001 **
	Moderate	26	65.00	
	Weak	8	20.00	
Age (year)	Mean ±SE	530 ±0.31		---
** (P<0.01).				

Two primary characteristics of autism spectrum disorders are limited interests and repetitive activities, as well as deficiencies in social interaction and communication. Patients may exhibit signs of ASD as early as 12 months, however it may take until later in childhood for these symptoms to be officially diagnosed as ASD (Tachibana et al., 2017). According to research on ASD, some patients have trouble updating links between unpleasant unconditioned stimuli and environmental signals (South et al., 2012). Lee and Odom (1996) study comparing the social interaction of two children, one with ASD and the other with a mental disability, who were both between the ages of 7 and 8. We watched how they interacted with their peers. The study's findings demonstrated that compared to other mentally challenged children, the ASD youngster was more reclusive while interacting socially with classmates.

The symptoms of ASD are defined in the following ways: anxiety related to language and social skills, variations in how the senses react to stimuli, Incapacity to interact with others and lack of speech and communication, even though linguistic skills are present (American Psychiatric Association, 2013). Out of 80 cases, Kadhim et al. (2015) reported: There were 40 children with ASD, 20 children without ASD (siblings), and 20 healthy children without relations. The risk of ASD males was larger in 31 (77.5%) than in ASD females (9 (22.5%)), with a 4:1 ratio. The previous study involved a mouse model in which a combination of the major isoforms (Shank3E13) was deleted, which exhibited behavioral defects including excessive grooming, decreased social interaction, rearing deficiency, locomotor and learning and memory impairment (Jaramillo et al., 2017). Other studies besides these have reported the same in similar Shank3 mouse models along with identifying reduced hippocampal long-term potentiation, morphologic and structural abnormalities including reduced spine density and increased dendritic length as well as the loss of synaptic proteins that associate with SHANK3 (Yang et al., 2012; Kouser et al., 2013).

### 3.2. Molecular Study:

Since the blood was processed right away, total RNA was successfully recovered from every sample. The range of total RNA concentration and purity was 65–98 ng/ $\mu$ l and 1.68–1.97 ng/ $\mu$ l, respectively. Following the RNA extraction procedures, cDNA reverse transcription was performed. Random hexamer primers that cover all RNA regions to produce cDNA were included in the kit.

#### 3.2.1. Quantitative Real-Time PCR Results:

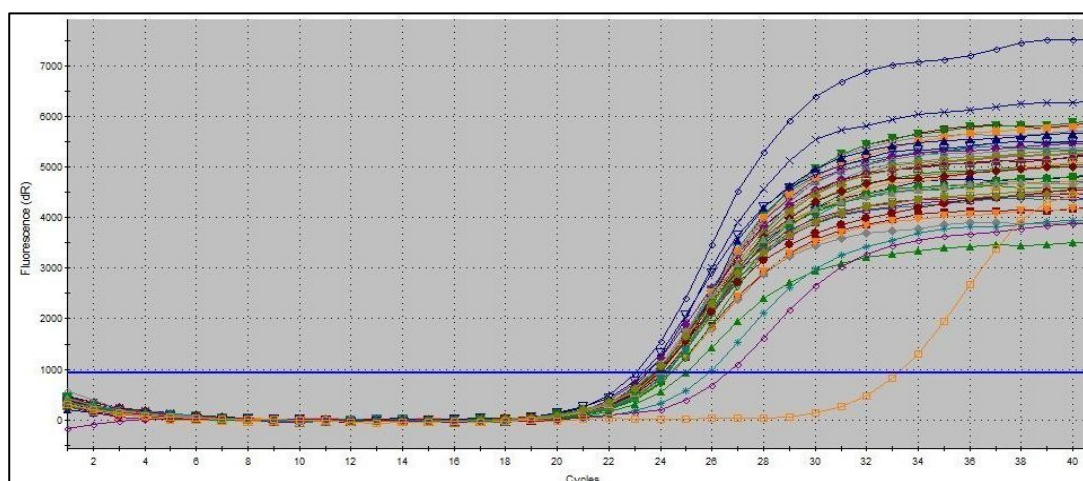
In this experiment, the EVA green was used for real-time PCR quantification. Any double-stranded DNA, including cDNA, was detected by the fluorescent dye, and the amplification was quantified as a Ct value. Higher copies of the target are present when the Ct value is smaller, and vice versa. A low

level of gene expression is indicated by high Ct values. The Ct value of gapdh did not significantly differ between the individuals and the healthy control group ( $1 \pm 0.00$ ). According to Table (2) and Figure (1), the patient groups' mean fold of gapdh gene expression was  $0.98 \pm 0.08$ .

**Table (2): Comparison of gapdh fold between study groups depending on  $2^{-\Delta Ct}$  Method.**

Group	Mean Ct of gapdh	$2^{-\Delta Ct}$	Expression group/ control group	mean fold of gapdh expression
Patients	29.977	9.5 E10	9.5 E10/9.7 E10	$0.98 \pm 0.08$
Control	29.946	9.7 E10	9.7 E10/9.7 E10	$1 \pm 0.00$
LSD value	---	---	---	0.217 NS

NS: Non-Significant.



**Figure (1): gapdh genes amplification plots by qPCR .Ct values was ranged from 23.32 to 25.4.The photograph was taken directly from Qtower2.0/2.2**

The application of housekeeping genes in molecular research is based on the premise that the cells are always active in the expression of the housekeeping genes (Reboucas, 2013). Gapdh is considered as one of the housekeeping genes most frequently utilized according to the gene expression data (Barber et al., 2005). Robert et al. (2005) also examined the expression of 1718 genes using qRT-PCR. They performed a gapdh gene as a reference gene in seventy two varieties of normal human tissues. When applied in clinical studies, they found that gapdh is a highly reliable protocol in normalizing qRT-PCR.

### 3.2.2. *Shank3* Gene Expression:

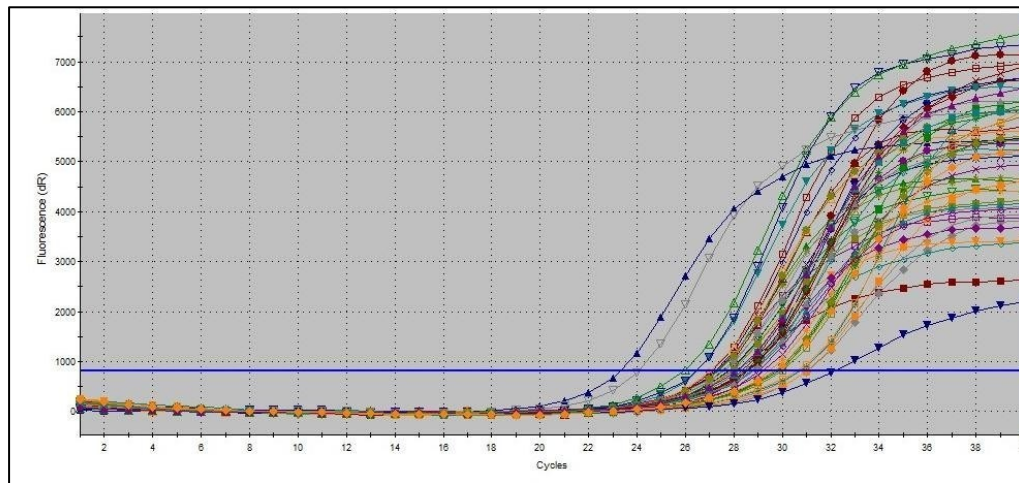
Expression of the *Shank3* gene was a significant decreased ( $0.374 \pm 0.18$ ) in ASD patients when compared to the healthy control group ( $1.00 \pm 0.00$ ) as shown in table (3), Figure (2).

**Table (3): Comparison of *Shank3* gene fold between groups depending on  $2^{-\Delta Ct}$  Method.**

Group	Mean Ct of <i>Shank3</i> gene	Mean Ct of gapdh gene	$\Delta Ct$	$\Delta\Delta Ct$	<i>Shank3</i> Fold change
Patients	20.01	20.94	-0.9379	-2.3461	$0.421 \pm 0.19$
Control	19.58	18.16	1.5256	1.5256	$1.00 \pm 0.00$
T-test	--	--	--	--	0.518 *
P-value	--	--	--	--	0.0476

\* ( $P \leq 0.05$ ).





**Figure (2): *Shank3* gene amplification plots by qPCR .Ct values was ranged from 28.34 to 32.13.The photograph was taken directly from Qtower2.0/2.2,**

The immense silencing of the genes due to gene mutations, especially *Shank3* mutations in three families were studied by Durand et al. (2007) who discovered that the frame shifts and translocations were associated to ASD. The mutations of *Shank3* can be found in approximately 0.5-1 percent of all individuals with ASD (Jiang and Ehlers, 2013). Those patients with autism and speech delay who were examined by Boccuto et al. (2013) found three cases of deletion of amino acids that received a frameshift mutation in *Shank3* and single case of insertion of an early stop codon. *Shank3* is one of the most researched genes that have been associated with ASD. Genetic studies carried out on patients with ASD have identified over 40 known mutations in the *Shank3* gene either leading to truncating variations or expected to be dangerous. (Leblond et al., 2014).

As it was demonstrated previously, mice targeted with the disruption of the PDZ domain of *Shank3* gene exhibited several behavioral abnormalities (Jaramillo et al., 2017).

In a cohort of 427 individuals with autism spectrum disorder (ASD), Gauthier et al. (2009) identified two *SHANK3* mutations, one deletion of one base pair (BP), was a de novo deletion that happened at an intronic splice site donor, and a one-base-pair (BP) missense mutation that was inherited by a father with epilepsy. The hypotheses are informed by a study of recurrent breakpoints in the *SHANK3* gene which result in the conclusion that defects in *SHANK3* most probably cause the autistic symptoms observed in some patients with 22q13 deletion syndrome (Bonaglia et al., 2006). One of the most described genomic rearrangements that are mostly linked to cognitive impairment is the 22q13 microdeletion syndrome that is linked to postnatal hypotonia, developmental delay, normal or accelerated somatic growth, non-development or severely retarded speech, autistic phenotype, and minor dysmorphic features.

### 3.3. Correlation between *Shank3* Expression and Ability to Speech, Education and social communication.

Table (4) showed there were a positive Correlation between *Shank3* gene expression to Ability to Speech, Ability to Education and Ability to Communication

**Table (4): Correlation between *Shank3* gene expression to Ability to Speech, Education and Communication**

Features	Correlation coefficient-r with fold of <i>Shank3</i> gene	P-value
<b>Ability to Speech</b>	0.010 *	0.953
<b>Ability to Education</b>	0.147 *	0.400
<b>Ability to social Communication</b>	0.314 *	0.070
* (P≤0.05) , NS: Non-Significant.		



Peca et al. (2011) created a model of *SHANK3* knockout mouse, in which a mutation in the PDZ domain created a range of behavior disturbances, such as deficiency of the social interaction, increased anxiety, and a set of resounding degrees of repetitive behaviors. Later on, Rendall et al. (2017) has reported that the knockout model shows an extended grooming period, augmented aggression, cognition, and better discrimination in pitch, which is a common phenomenon in persons with ASD (Remington and Fairnie, 2017). In a second study, Mei et al. (2016), showed that restoring expression of *SHANK3* in adult knockout mice is effective to normalize repetitive grooming and social behavior, indicating the role of this gene to produce modulation in behavioral phenotype even in later life.

Together, results in different *SHANK3* mouse models indicate that defect in the gene may affect features related to speaking as well as bipolar disorder and autism spectrum disorder (ASD). Both ASD and PhelanMcDermid syndrome (PMS) are characterized by speech and language issues, such as the absence or delayed development of the speech (and the use of simplified language), as well as their impairment (De Rubeis et al., 2018). According to these findings, it is postulated that PMS is an essentially a language, cognitive disorder with different neurophysiological processes being involved, unlike idiopathic ASD (Ponson et al., 2018). However, shared symptoms including engaged interests and repetitive therapy sustain in the two disorders (Soorya et al., 2013).

There is an increasing number of studies that confirm the correlation of *SHANK3* disruption with neurodevelopmental and psychiatric diseases and conditions, such as ASD and schizophrenia (De Sena Cortabitarte et al., 2017). As an example, de novo mutation in *SHANK3* was found in one person, and there were nine inherited nonsynonymous mutations found in *SHANK3* out of 400 persons having autism (Moessner et al., 2007). On the same note, a de novo splice-site deletion and a heritable missense mutation of *SHANK3* was also reported in relation to ASD by Gauthier et al. (2010). In a different study, a genetic analysis undertaken by Waga et al. (2011) using 128 individuals with ASD revealed multiple variants of the *SHANK3* gene as a six-amino-acid deletion upstream of the SH3 domain, missense mutation (arginine to histidine at position 656) within the PDZ domain, and a 10-basepair GC insertion/deletion polymorphism at 9 bp downstream of the 3 end of exon 11. The variants were not identified in 228 control subjects and the alterations were related with a harsh speech development.

### Conclusions:

This study confirms the *Shank3* gene that is an important causable gene for the ASD and related to the autistic features. The evidence from previous studies has shown the different type of *Shank3* gene mutation, but this study concludes the down-regulation of *Shank3* is associated to Speech, Education and Social Communication ability. The results of recent study it may be promote the therapeutic approaches for the treatment of ASD, such as restoration of normal *Shank3* gene expression.

### References:

1. Tomchek SD, Dunn W. Sensory processing in children with and without autism: a comparative study using the short sensory profile. *Am J Occup Ther.* 2007;61(2): 190–200.
2. Moore DJ. Acute pain experience in individuals with autism spectrum disorders: a review. *Autism.* 2015;19:387-399.
3. Phelan K, McDermid HE (2012) The 22q13.3 Deletion Syndrome (PhelanMcDermid Syndrome). *Mol Syndromol* 2: 186–201.
4. Mitchell, K.J. (2010) The genetics of neurodevelopmental disease. *Curr. Opin. Neurobiol.* 21, 1–7
5. Geschwind DH (2009) Advances in autism. *Annu Rev Med* 60: 367–380.
6. Huguet G, Ey E, Bourgeron T (2013) The Genetic Landscapes of Autism Spectrum Disorders. *Annu Rev Genomics Hum Genet* 14: 191–213.





7. Sudhof TC (2008) Neuroligins and neuexins link synaptic function to cognitive disease. *Nature* 455: 903–911
8. Toro R, Konyukh M, Delorme R, Leblond C, Chaste P, et al. (2010) Key role for gene dosage and synaptic homeostasis in autism spectrum disorders. *Trends Genet* 26: 363–372
9. Leblond.C.L, Nava.C, Polge.A, Gauthier.J, Guillaume Huguet,G. et al., : Meta-analysis of SHANK Mutations in Autism Spectrum Disorders: A Gradient of Severity in Cognitive Impairments. *PLOS genetics*.2014;10(9): 1-15
10. Livak, K.J. and Schmittgen, T.D. ( 2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$  CT Method. *Methods*. 25, 402–408
11. SAS. (2004). SAS / STAT Users Guide for Personal Computers. Release 7.0. SAS Institute Inc., Cary, NC., USA. (SAS = Statistical Analysis System).
12. Yang M, Bozdagi O, Scattoni ML, Wöhr M, Rouillet FI, Katz AM, et al., (2012) Reduced excitatory neurotransmission and mild autism-relevant phenotypes in adolescent Shank3 null mutant mice. *J Neurosci* 32:6525–6541
13. Kouser M, Speed HE, Dewey CM, Reimers JM, Widman AJ, Gupta N, Liu S, et al., (2013) Loss of predominant Shank3 isoforms results in hippocampus-dependent impairments in behavior and synaptic transmission. *J Neurosci* 33:18448–18468.
14. South M, Newton T, Chamberlain PD (2012).Delayed reversal learning and association with repetitive behavior in autism spectrum disorders. *Autism Res* 5:398–406.
15. Leblond CS, Nava C, Polge A, Gauthier J, Huguet G, Lumbroso S, Giuliano F, et al., (2014) Meta-analysis of SHANK mutations in autism spectrum disorders: a gradient of severity in cognitive impairments. *PLoS Genet* 10:e1004580
16. Jaramillo TC, Speed HE, Xuan Z, Reimers JM, Escamilla CO, Weaver TP,et al., (2017) Novel Shank3 mutant exhibits behaviors with face validity for autism and altered striatal and hippocampal function. *Autism Res* 10:42–65.
17. Jiang YH, Ehlers MD (2013). Modeling autism by SHANK gene mutations in mice. *Neuron* 78:8–27.
18. Tachibana Y, Miyazaki C, Ota E, Mori R, Hwang Y, Kobayashi E, Terasaka A, Tang J, Kamio Y (2017) A systematic review and meta-analysis of comprehensive interventions for pre-school children with autism spectrum disorder. *PLoS One* 12: e0186502
19. Mitz AR, Philyaw TJ, Boccuto L, Shcheglovitov A, Sarasua SM, Kaufmann WE, Thurm A (2018) Identification of 22q13 genes most likely to contribute to Phelan McDermid syndrome. *Eur J Hum Genet* 26:293–302
20. Bonaglia, M. C., Giorda, R., Mani, E., Aceti, G., Anderlid, B. M., Baroncini, A.,et al., (2006). Identification of a recurrent breakpoint within the SHANK3 gene in the 22q13.3 deletion syndrome. *J.Med.Genet.* 43, 822-828.
21. Kadhim,B. Al-Kazaz, Abdul-Kareem and and Al. Deresawi,M.S. (2015). Molecular Genetics Study on Autostatic patients in iraq. *Iraq Journal of Science*.56(1).
22. Gauthier, J., Bonnel, A., St Onge, J., Karemera, L., Laurent, S., Mottron, L., et al., (2005). NLGN3/NLGN4 gene mutations are not responsible for autism in the Quebec population. *Am.J.Med.Genet.B Neuropsychiatr.Genet.* 132, 74-75.
23. Reboucas E.; Costa J.; Passos M.; Passos J.; Hurk R. and Silva J. (2013). Real Time PCR and Importance of Housekeeping's Genes for Normalization and Quantification of mRNA Expression in Different Tissues. *Brazil Arch Biol Technol.* 56: 143-154



24. Barber D. (2005). GAPDH as a housekeeping gene: analysis of GAPDH mRNA expression in a panel of 72 human tissues. *Physiological Genomics*; vol. 21 (3): 389-395.
25. Robert B.; Harmer W.; Coleman A. and Clark B. (2005). GAPDH as a housekeeping gene: analysis of GAPDH mRNA expression in a panel of 72 human tissues. *Physiol Genom.* 21: 389– 395.
26. American Psychiatric Association (2013). *Diagnostic and statistical manual of mental disorders*. 5th ed. Arlington, VA: American Psychiatric Association.
27. Pérez E, Acero-Ferrero M and Herrero ML.(2019). Improvement of Planning Skills in Children With Autism Spectrum Disorder After an Educational Intervention: A Study From a Mixed Methods Approach. *Front Psychol.* 17;10:2824.
28. Lee, S., and Odom, S.L. (1996). The Relationship between Stereotypic Behavior and Peer Social Interaction for Children with Severe Disabilities. *Research and Practice for Persons with Severe Disabilities*, 21, 88 - 95.
29. De Rubeis S., Siper P. M., Durkin A., et al.(2018). Delineation of the genetic and clinical spectrum of Phelan-McDermid syndrome caused by SHANK3 point mutations. *Molecular Autism.* 9(1):1–20.
30. Durand C. M., Betancur C., Boeckers T. M., et al.(2007). Mutations in the gene encoding the synaptic scaffolding protein SHANK3 are associated with autism spectrum disorders. *Nature Genetics.* (1):25–27.
31. Mei Y., Monteiro P., Zhou Y., et al.(2016). Adult restoration of Shank3 expression rescues selective autistic-like phenotypes. *Nature.* 530(7591):481–484.
32. Ponson L., Gomot M., Blanc R., et al. (2018). 22q13 deletion syndrome: communication disorder or autism? Evidence from a specific clinical and neurophysiological phenotype. *Translational Psychiatry.* 8(1)
33. Gauthier J., Champagne N., Lafrenière R. G., et al.(2010). De novo mutations in the gene encoding the synaptic scaffolding protein SHANK3 in patients ascertained for schizophrenia. *Proceedings of the National Academy of Sciences.* 107(17):7863–7868.
34. Moessner R., Marshall C. R., Sutcliffe J. S., et al(2007). Contribution of SHANK3 mutations to autism spectrum disorder. *The American Journal of Human Genetics.*;81(6):1289–1297.
35. Soorya L., Klevzon A., Zweifach J., et al.(2013). Prospective investigation of autism and genotype-phenotype correlations in 22q13 deletion syndrome and SHANK3 deficiency. *Molecular Autism.* 4(1):18–17.
36. Boccuto L., Lauri M., Sarasua S. M., et al.(2013). Prevalence of SHANK3 variants in patients with different subtypes of autism spectrum disorders. *European Journal of Human Genetics.* 21(3):310–316.
37. Peça J., Feliciano C., Ting J. T., et al.(2011). Shank3 mutant mice display autistic-like behaviours and striatal dysfunction. *Nature.* 2011;472(7344):437–442.
38. Rendall A. R., Perrino P. A., Buscarello A. N., Fitch R. H. (2019). Shank3B mutant mice display pitch discrimination enhancements and learning deficits. *International Journal of Developmental Neuroscience.* 72(1):13–21.
39. Remington A., Fairnie J. A sound advantage: increased auditory capacity in autism. *Cognition.* 2017;166:459–465..
40. de Sena Cortabitarte A., Degenhardt F., Strohmaier J., et al.(2017). Investigation of SHANK3 in schizophrenia. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics.* 174(4):390–398.





41. Waga, Chikakoa; Okamoto, Nobuhikoc; Ondo, Yumikoa; Fukumura-Kato, Reikoa; et al.,( 2011). Novel variants of the SHANK3 gene in Japanese autistic patients with severe delayed speech development. *Psychiatric Genetics* 21(4):p 208-211,
42. Reboucas E.; Costa J.; Passos M.; Passos J.; Hurk R. and Silva J. (2013). Real Time PCR and Importance of Housekeepings Genes for Normalization and Quantification of mRNA Expression in Different Tissues. *Brazil Arch Biol Technol.* 56: 143-154.
43. Barber D. (2005). GAPDH as a housekeeping gene: analysis of GAPDH mRNA expression in a panel of 72 human tissues. *Physiological Genomics*; vol. 21 (3): 389-395.
44. Robert B.; Harmer W.; Coleman A. and Clark B. (2005). GAPDH as a housekeeping gene: analysis of GAPDH mRNA expression in a panel of 72 human tissues. *Physiol Genom.* 21: 389– 395.